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Cytotoxic cuparene sesquiterpenes from Laurencia microcladia

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Abstract—Three new cuparene sesquiterpenes 1–3 were isolated from the organic extract of the red alga *Laurencia microcladia*, collected at Chios island in the North Aegean Sea, Greece. The structures and the relative stereochemistry of the compounds are proposed on the basis of their spectral data. Metabolite 2 shows an unprecedented (for the cuparene class of sesquiterpenes) migration of the C-1 methyl group. All metabolites 1–3 were found to exhibit significant cytotoxic activity against two lung cancer cell lines. © 2005 Elsevier Ltd. All rights reserved.

The red algae belonging to the genus *Laurencia* (Ceramiales, Rhodomelaceae) have been proven to be rich sources of secondary metabolites. The majority of these metabolites are characterized by their relatively high degree of halogenation. Their ecological roles have not been clearly elucidated, but it is suggested that they function as chemical defenses against marine herbivores. Despite the fact that *Laurencia* has been studied extensively with respect to secondary metabolite chemistry, new studies on members of this genus frequently lead to the isolation of novel intriguing structures, especially terpenoids and C₁₅-acetogenins. Structures,

As part of our ongoing program aimed at the isolation of biologically active compounds from marine organisms of the Greek seas, 8-10 we investigated the secondary metabolite content of the red alga *Laurencia microcladia*, collected from the south coast of Chios island. We herein describe the isolation and structure elucidation of the new sesquiterpenes 1–3.

The collected alga specimens were initially freeze-dried (291.4 g dry weight) and then exhaustively extracted with mixtures of CH₂Cl₂/MeOH (3:1) at room temperature. The extract was concentrated to give a dark green

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residue (12.2 g), which was subjected to vacuum column chromatography (VCC) on silica gel, using cyclohexane with increasing amounts (10%) of EtOAc as the mobile phase. Following a combination of chromatographic techniques, including VCC, gravity column and finally HPLC purifications, compounds 1 (4.3 mg), 2 (3.0 mg), and 3 (1.5 mg) were isolated in pure form.

Metabolite 1, a bromo sesquiterpene ether, was isolated as a colorless oil, $[\alpha]_D^{20}$ –29.00 (c 0.1, CH₂Cl₂). The HRFAB-MS measurements supported the molecular formula $C_{15}H_{19}OBr$ (*m/z* 294.0611 [M⁺]) and the M⁺ peaks in the EI-MS spectrum, at m/z 294 and 296 with relative intensities 1/1, revealed the presence of one bromine atom in the molecule. The presence of a substituted benzene ring was evident from the UV spectrum, 11 which had maxima at 275 and 285 nm and the IR spectrum with absorbances at 1648 and 1508 cm⁻¹. The IR absorption at 1241 cm⁻¹ and the absence of an absorption band for hydroxyl or carbonyl groups indicated that the oxygen atom was involved in an ether linkage. The ¹³C NMR spectrum of 1 (Table 1) showed signals for 15 carbons with the multiplicities of the carbon signals determined from the DEPT spectrum as: six quaternary, three methine, two methylene, and four methyl carbon atoms. The ¹H and ¹³C NMR spectra of 1 confirmed the presence of the aromatic ring (δ 6.56/117.6, 7.18/127.8). Since 1 has six degrees of unsaturation, with no other additional double bonds, it must contain two other rings, one of which is the ether-containing ring

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Table 1. ¹H and ¹³C NMR data (400 and 50.3 MHz, respectively) for metabolites 1-3

		Metabolite 1	olite 1			Metabolite 2	2		Me	Metabolite 3		
	(ϱ) \mathbf{H}_{1}	13C (δ)	HMBC		(ϱ) H_1	13C (δ)	HMBC		(δ) H ¹	13C (δ)	HMBC	
C-1		45.9	I	C-1		147.1		C-1		54.3		
C-2	1	41.2		C-2	1	49.3	1	C-2	1	137.1	1	
H-3	4.10 (d, 5.1)	86.3		H-3	2.08 (m)	44.8	1	C-3	1	131.8	1	
H-4	2.09 (m)	30.0		H-4	2.53 (m) 2.06 (m)	38.3	C-2	H-4	1.93 (m) 2.29 (m)	35.7	1	
H-5	1.88 (m)	41.3		H-5	5.75 (br s)	131.1	C-2, C-3, C-4	H-5	1.38 (m) 1.72 (m)	41.2		
C-6		133.1		C-6		123.1		C-6		148.6	1	
C-7		151.8		C-7		152.1		H-7	6.62 (s)	112.2	C-11	
8-H	6.56 (s)	117.6	C-6, C-7, C-10, C-15	H-8	6.82 (s)	117.0	C-6, C-10, C-15	C-8	:	153.4	1	
C-9	1	136.3		C-9	1	137.9		C-9	1	130.4	1	
C-10		114.5		C-10	1	114.0	1	H-10	7.00 (d, 7.8)	129.0	C-6, C-8, C-15	
H-11	7.18 (s)	127.8		H-111	7.17 (s)	131.9	C-1, C-7, C-9	H-11	6.71 (d, 7.8)	118.7	C-7	
H-12	(s) 68·0	18.1		H-12	0.87 (s)	20.5	C-1, C-2, C-3, C-13	H-12	1.35 (s)	10.1	C-2, C-3	
H-13	0.97 (s)	20.4		H-13	0.97 (s)	26.1	C-1, C-2, C-3, C-12	H-13	1.68 (s)	14.3	C-2, C-3, C-4	
H-14	1.22 (s)	14.9		H-14	1.01 (d, 6.8)	14.2	C-3, C-4	H-14	1.35 (s)	26.7	C-1, C-5, C-6	
H-15	2.26 (s)	22.4	C-8, C-9, C-10	H-15	2.32 (s)	22.7	C-8, C-9, C-10	H-15	2.19 (s)	14.9	C-9, C-8	
				OH-C-7	5.33 (s)		C-7, C-8	OH-C-8	4.63 (s)			

Chemical shifts are expressed in ppm. J values in parentheses are in Hz.

and the other a five-membered carbocyclic ring. Signals corresponding to four tertiary methyl groups appeared at δ 0.89/18.1, 0.97/20.4, 1.22/14.9 and 2.26/22.4. A doublet at δ 4.10 was attributed to a proton on carbon C-3 $(\delta 86.3)$, participating in the ether bridge. The ether bridge was placed on carbons C-7 and C-3 because of the heteronuclear correlation between H-3 (δ 4.10) and C-7 (δ 151.8). The correlation of signals at δ 0.89 (H-12)/0.97 (H-13) with those at 45.9 (C-1), 41.2 (C-2), and 86.3 (C-3) confirmed the position of the gem-dimethyl groups on C-2. The relative stereochemistry of 1 was assigned on the basis of NOESY experiments. The strong NOE correlations between H-14/H-13 and between H-13/H-3 determined the stereochemistry at C-1 and C-3. The ¹H and ¹³C NMR data of compound 1 are in good agreement with reported values for the debromo-analogue 4 isolated previously from Laurencia okamurai. 12 In view of the above-mentioned data and considering the biosynthetic relations, the proposed structure for ether metabolite 1 is shown in Figure 1.

Metabolite 2, a bromo sesquiterpene phenol, obtained as a colorless oil with $\left[\alpha\right]_{D}^{20}$ +5.00 (c 0.06, CH₂Cl₂), showed LREI-MS signals at m/z 294/296 [M⁺], with relative intensities 1/1, suggesting the presence of one bromine atom. The HRFAB-MS measurements supported the molecular formula $C_{15}H_{19}OBr$ (m/z 294.0628 [M⁺]). The presence of a benzene ring was evident from the IR spectrum, which showed absorbances at 1642 and $1465 \,\mathrm{cm}^{-1}$. The intense sharp absorptions at v_{max} 3428 and 1157 cm⁻¹ indicated the presence of a hydroxyl functionality in the molecule. ¹³ The ¹³C NMR spectrum of 2 (Table 1) showed signals for 15 carbons. Multiplicities for the carbon signals were determined from the DEPT spectra as: six quaternary, four methine, one methylene and four methyl carbons. The ¹H and ¹³C NMR spectra displayed resonances for a secondary methyl (δ 1.01 d/14.2), one aromatic methyl (δ 2.32/ 22.7), two quaternary methyls (δ 0.87/20.5 and δ 0.97/ 26.1), an olefinic proton (δ 5.75/131.1), two aromatic protons (δ 6.82/117.0 and δ 7.17/131.9), one methine (δ 2.08/44.8), one methylene (δ 2.53, 2.06/38.3), and one exchangeable proton (δ 5.33). The position of the olefinic proton of the trisubstituted double bond at C-5 was determined from correlations between H-5/C-2, H-5/C-3, and H-5/C-4. Moreover, the correlation of the vinyl carbon (δ 147.1) with the aromatic proton H-11 (δ 7.17) as observed in the HMBC spectrum confirmed the position of the double bond between C-1 and C-5. The correlation of signals at δ 0.87 (H-12)/0.97 (H-13) with 147.1 (C-1), 49.3 (C-2), and 44.8 (C-3) determined the position of the gem-dimethyl groups on C-2, vicinal to the secondary methyl, located on C-3. Moreover, NOE correlations between the aromatic proton H-11 and the geminal methyls C-12, C-13 further supported the position of the methyls (C-12 and C-13) on C-2. Combination of these data led to the assignment of the structure as shown in Figure 1. To the best of our knowledge, this is the first report of a cuparene sesquiterpene with a double bond between C-1 and C-5 resulting from methyl migration on either the C-2 or C-3 position. Metabolite 2 could have potentially arisen from rearrangement of compound 1, but the fact that

Figure 1. Structures of metabolites 1-5.

no conversion or instability of metabolite 1 was observed during the chromatographic separations supports 2 as an algal secondary metabolite.

Metabolite 3 was isolated as a colorless oil, with $[\alpha]_D^{20}$ +40.00 (c 0.07, CH₂Cl₂). The HRFAB-MS measurements supported the molecular formula $C_{15}H_{19}O$ (m/z 215.1443 [M-1]^+). The presence of a substituted benzene ring was suggested by the UV spectrum, 14 which had maxima at 274 and 280 nm. The IR absorptions at $v_{\rm max}$ 3422 and 1639 cm⁻¹ were attributed to a hydroxyl and a double bond, respectively. The ¹³C NMR spectrum displayed fifteen signals (Table 1) and their multiplicities were determined from the DEPT spectra as: six quaternary (two olefinic, three aromatic, and one aliphatic), three methines, two methylenes, and four methyl carbons. The ¹H NMR spectrum exhibited absorptions at δ 6.62 (s, 1H), 6.71 (d, 1H), and 7.00 (d, 1H) due to aromatic protons, an aromatic methyl group at 2.19 (s, 3H), a tertiary methyl group at δ 1.35, and two vinyl methyls at δ 1.35 and 1.68. The vinyl methyl at δ 1.35 appears unexpectedly at high field frequency probably due to the magnetic anisotropy exerted by the benzene ring. 15 Moreover, the lack of any NOE between H-7/H-11 and the methyl group at δ 1.68 supported its position on C-13. Comparison of the NMR data of 3 with reported values for isolaurene 5 was in accordance with 3 being its hydroxyl derivative. 16,17 The chemical shift of the aromatic methyl group at δ 2.19/14.9 at higher field compared to isolaurene (δ 2.30/20.9) supported the position of the hydroxyl functionality on C-8. Furthermore, an HMBC correlation between H-15 (δ 2.19) and C-8 (δ 153.4) further confirmed the position of the hydroxyl group. In view of the above-mentioned data, the proposed structure for metabolite 3 is shown in Figure 1.

The cytotoxicities of compounds 1–3 were assayed against NSCLC-N6 and A549 lung cancer cell lines. Metabolite 1 showed moderate cytotoxicity: IC $_{50}$ = 196.9 and 242.8 μ M against NSCLC-N6 and A549 cancer cell lines, respectively. Metabolites 2 and 3 showed stronger levels of activity with: IC $_{50}$ = 73.4 μ M (NSCLC-N6) and 52.4 μ M (A549) and IC $_{50}$ = 83.7 μ M (NSCLC-N6) and 81.0 μ M (A549), respectively. The higher cytotoxicity of metabolites 2 and 3 could be attributed to the phenolic hydroxyl and/or the presence

of the double bond in the five-membered ring. The presence of bromine in these molecules does not seem to affect significantly their activity according to our results.

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- 11. Spectral data of compound 1: $[\alpha]_D^{20} 29.00$ (c 0.1, CH₂Cl₂); HRFAB-MS: m/z [M]⁺; 294.0611 (294.0619 calculated for C₁₅H₁₉O⁷⁹Br); IR (KBr): $\nu_{\rm max}$ 1648, 1541, 1508, 1391, 1376, 1241, 1162, 1041, 812 cm⁻¹; UV $\lambda_{\rm max}^{\rm CH₂Cl₂}$ (nm) (log ε) 237 (3.02), 275 (2.42), 285 (2.56), 295 (2.44). NMR data are shown in Table 1.
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- 13. Spectral data of compound 2: $[\alpha]_D^{20} + 5.00$ (c 0.06, CH₂Cl₂); HRFAB-MS: m/z [M]⁺; 294.0628 (294.0619 calculated for C₁₅H₁₉O⁷⁹Br); IR (KBr): v_{max} 3428, 2952, 2921, 2861, 1642, 1465, 1157 cm⁻¹; UV $\lambda_{max}^{CH_2Cl_2}$ nm (log ε) 237 (3.19) 290 (2.92) NMR data are shown in Table 1
- 237 (3.19), 290 (2.92). NMR data are shown in Table 1.

 14. Spectral data of compound 3: $[\alpha]_D^{20}$ +40.00 (*c* 0.07, CH₂Cl₂); HRFAB-MS: m/z [M-1]⁺; 215.1443 (215.1437 calculated for C₁₅H₁₉O); IR (KBr): ν_{max} 3422, 1639,

- 725 cm⁻¹; UV λ_{max}^{CH₂Cl₂} nm (log ε) 235 (2.88), 274 (2.67), 280 (2.55). NMR data are shown in Table 1.
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